

Action D1.
Analyses of data and samples, evaluation
and optimisation of techniques

Deliverable:

List of fungi associated to *Xylosandrus* spp.

1

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Summary

Insects, like many other organisms, live in association with many fungal symbionts, which can have a positive (i.e., mutualistic), negative (i.e., parasitic) or neutral (i.e., commensalistic) impact on their host's fitness. Symbiotic fungi associated with Ambrosia beetles contribute with insect damage to the impact to invaded environments. Also of providing food for the insect development stages, some of the Ambrosia beetle symbiotic fungi are severe pathogens of plant hosts. The most relevant example is Xyleborus glabratus carrying the fungus Raffaela lauricola cause of lethal vascular wilt of avocado, that is devastating the plantations in South-eastern United States. Thus, monitoring of alien fungi introduced through specific pathways (mostly trading of living plants), their identification and determination of pathogenicity behaviour is essential to design and apply prevention and mitigation quarantine measures. This deliverable describes the fungal community isolated from X. compactus and X. crassiusculus.

1. Bark beetle' and obligate symbiosis with fungi

Bark beetle is both a taxonomic and ecological designation. In the taxonomic sense, bark beetles are all species in the weevil subfamily Scolytinae, including species that do not consume bark. In the ecological sense, bark beetles are species of Scolytinae whose larvae and adults live in and consume phloem of trees and other woody plants. They are not obligatory associated with fungal symbionts, while ambrosia beetles are obligately associated with nutritional fungal symbionts. Obligate symbiosis with fungi is present in at least 11 independent scolytinae and platypodine groups. Ambrosia beetles are therefore not monophyletic, and the name is not a taxonomic designation. Ambrosia beetles are derived from bark beetles (Coleoptera: Curculionidae: Scolytinae). Bark beetles colonize and consume phloem, a tissue that is more nutrient-rich than wood. Bark beetles, like ambrosia beetles, are also often associated with fungal symbionts, usually ascomycotan and rarely basidiomycotan fungi, and the intensity of association is more variable, ranging from facultative to obligate (You L, 2015).

One of the most common symbioses in any forest ecosystem occurs between wood-boring insects and fungi.

This kind of symbiosis between Ambrosia beetles (Coleoptera: Curculionidae: Scolytinae and Platypodinae) and ambrosia fungi is ideal for studying many symbiosis-related questions. One reason is the diversity of ambrosia beetles which represent about 3000 species of wood-boring weevils that repeatedly evolved obligate symbioses with nutritional fungi possibly up to 16 times. Another reason is the easy manipulation of the symbiosis. Although the two partners require each other to complete their life cycle, they are perfectly separable in vitro. The beetles' transport-specific fungal symbionts from their natal galleries to newly established galleries in a storage organ termed a mycangium, but both can be kept in the laboratory on artificial media. Furthermore, their importance needs to be taken into consideration as in recent years, several ambrosia beetle-fungus symbioses have developed outbreaks causing significant economic and ecological damages. Unquestionably, it is important to understand the interactions between the beetle and the fungus has immediate scientific, economic and ecological implications (You Li1, 2018). Moreover, like many insects, ambrosia beetles may carry commensalist fungi on their body.

2. *Xylosandrus crassiusculus* (Motschulsky)

Xylosandrus crassiusculus (Asian ambrosia beetle or granulate ambrosia beetle) it is a highly polyphagous pest of woody plants of Asian origine and has been spread most probably with trade of plants and wood. In Africa it arrived hundreds of years ago while in the last few years it has been introduced to at least 14 African countries, 25 USA states, 3 countries of Central America, 2 South American countries, 6 countries of Oceania and 2 European countries (EPPO 2015, Fletchmann and Atkinson 2016), Italy and France, recently adding Spain (Gallego et al., 2017). Since the 1970s it has become a pest of fruit tree orchards and ornamental tree nurseries in the USA (EPPO Alert-list). It was first found in Europe in 2003, in cross-vane traps set up in Tuscany (Livorno, NW Italy) where no specific control measures were adopted (Pennacchio et al. 2003; EPPO 2015). Later, carob trees attacked by *X. crassiusculus* were found in in orchards in Central-North Italy and in gardens of nearby Liguria (Alassio and Pietraligure, NW Italy) in 2007 and 2008, and in NE Italy in Veneto (EPPO 2015) and Friuli Venezia Giulia (2015, personal observation of Massimo Faccoli). Maybe via Liguria, in 2014 the species arrived in SE France and in the Spanish Valencia Region.

Adults are small dark reddish brown scolytids (female: 2-3 mm long, males: 1.5 mm). Larvae are white, legless, C-shaped with a well-developed capsule, and cannot be easily distinguished from other scolytids. Populations essentially contain females (1:10 male-female ratio). Adult males do not fly and remain inside the galleries. Also *X. crassiusculus* is an inbreeding species (females mate with their brothers).

When females emerge, they leave infested plants and fly to new hosts. They start to bore a tunnel (round entrance hole of 2 mm diameter) with a brood chamber and one or more branches into the sapwood (and sometimes the heartwood). Eggs are laid in the brood chamber. Larvae have a length about 3.5 mm. and hatch and feed on the symbiotic fungus growing inside the galleries (Gardner, 1934, CABI Factsheet).

2.1. List of fungi isolated from *Xylosandrus crassiusculus*

The fungal isolation was done according to the protocol described on **SAMFIX Deliverable: Fast routine protocol for detection of fungal symbiotic community associated with trapped Xylosandrus**. Briefly, the insects are crumbled in PBS and serial dilutions are plated on PDA. Different colonies are subcultured in new PDA plates. Morphotype designations are confirmed by ITS DNA sequencing and compared with sequences available at NCBI database (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>). The fungi isolated from *X. crassiusculus* are on Table 1.

Table 1. List of fungi isolated from *Xylosandrus crassiusculus*

| SPECIES (best hit) | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|---|--------|--------------|-------------------------|-----------------------------|----------------------------------|--|---|
| <i>Penicillium expansum</i> (<i>P. spuinulosum</i>) | A | Eurotiales | abdomen, head, external | Plant pathogen/Saprotrophs | Cosmopolitan | Numerous hosts. Fruit, decaying vegetation, seeds, etc. Causing Blue mold, fruit rot, postharvest decay. | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Alternaria infectoria</i> | A | Pleosporales | external | Plant pathogen | Widespread in temperate regions. | Substrate: Kernels, leaves. Disease: Black point disease. | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Bipolaris sorokiniana</i> | A | Pleosporales | abdomen | Plant pathogen | Cosmopolitan. | Leaf spot, seedling blight, and root rot. Primarily on <i>Poaceae</i> but also numerous and diverse other hosts. | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Clonostachys rosea</i> | A | Hypocreales | mycangia | Plant pathogens/Saprotrophs | Cosmopolitan | Various plant parts both living and newly killed, associated with bark beetle galleries | Nygren et al., 2018 |
| <i>Cryphonectria parasitica</i> | A | Diaporthales | mycangia | Plant pathogen | Asia, Europe, North America | <i>Castanea</i> spp., <i>Fagus sylvatica</i> , <i>Quercus</i> spp. (Fagaceae). | https://nt.ars-grin.gov/fungaldat/abases |

| SPECIES (best hit) | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|--|--------|-----------------|-----------------------------------|-------------------------------|---|--|---|
| <i>Didymella glomerata/ D. fabae Aspergillus niger</i> | A | Pleosporales | external | Plant pathogens/Saprotrophs | Cosmopolitan | Various plant genera. Opportunistic pathogen. Found in association with blights, leaf spots, fruit rots. | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Geosmithia pallida</i> | A | Hypocreales | abdomen, mycangia, external | Plant pathogen | Widespread | Associated with bark beetle <i>Pseudopithyophthorus pubipennis</i> ; Hosts: <i>Quercus</i> spp. (Fagaceae), <i>Prunus</i> spp., <i>Malus</i> (Rosaceae); Causing Foamy bark canker, dieback, death | Kolarik et al., 2017 |
| <i>Pestalotiopsis vismiae</i> | A | Xylariales | mycangia | Plant pathogen | Asia (China, India), North America (USA). | Numerous hosts. Substrate: Trunk, bark, leaves, petioles, roots. Disease Note: Bark cracking, lesions; trunk disease. | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Acremonium roseolum</i> | A | Hypocreales | abdomen | Unassigned | Asia (Japan) , South America (Brasil) | <i>Cryptomeria japonica</i> and <i>Manihot esculenta</i> | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Cladosporium sp.</i> | A | Capnodiales | external | Saprotrophs | Cosmopolitan | Multiple genera in multiple families. Plant material and other organic substrates. Various spots and rots. | Bensch et al., 2012 |
| <i>Paraconiothyrium archidendri</i> | A | Xylariales | abdomen, head, mycangia, external | Plant pathogen | Asia (Myanmar) | Leaf spot on <i>Pithecellobium bigeminum</i> (Fabaceae). | Verkley et al., 2014 |
| <i>Pleurostoma richardsiae</i> | A | Calosphaeriales | external | Pathogen | Widespread | Wood streaking, canker; also a human pathogen. On <i>Olea europaea</i> (Oleaceae), <i>Vitis vinifera</i> (Vitaceae), <i>Prunus dulcis</i> (Rosaceae) and reports from diverse hosts. | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Trichoderma harzianum</i> | A | Hypocreales | external | Saprotrophs/Fungal antagonist | Cosmopolitan | Found on roots and other plant parts on numerous hosts; causing soft roots | Bissett et al., 2015; Han et al., 2017 |
| <i>Xenoacremonium falcatus</i> | A | Hypocreales | external | Unassigned | Asia, Europe | <i>Castanea sativa</i> ; other substrates | Aghyeva et al., 2017 |

| SPECIES (best hit) | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|-------------------------------------|--------|--------------|-----------------------------------|-------------------------------|--------------|--|---|
| <i>Fusarium solani</i> | A | Hypocreales | abdomen, head, mycangia, external | Plant pathogen | Cosmopolitan | Broad host range; associated with ambrosia beetles | Sharma and Marques, 2018 |
| <i>Pithomyces chartarum</i> | A | Pleosporales | head, external | Plant pathogens/Saprotrophs | Cosmopolitan | From <i>Pithomyces chartarum</i> leaves - Diverse plant families, but especially Poaceae. | https://nt.ars-grin.gov/fungaldat/abases |
| <i>Cladosporium cladosporioides</i> | A | Capnodiales | abdomen, head, external | Saprotrophs/hyperparasitic | Cosmopolitan | Multiple genera in multiple families; Leaves, seeds, inflorescences, often dead plant material; Causing leaf spot and blight, flower blight, scab, sooty mold. | Barkat et al., 2016 |
| <i>Talaromyces minioluteus</i> | A | Eurotiales | mycangia, external | Plant pathogens/Saprotrophs | Cosmopolitan | Post harvest fruit rot. | Palou et al., 2013 |
| <i>Alternaria alternata</i> | A | Pleosporales | abdomen, head, mycangia, external | Plant pathogens/Saprotrophs | Widespread | Wide-host range | Feng, Zheng, 2007 |
| <i>Trichoderma atroviride</i> | A | Hypocreales | abdomen, external | Saprotrophs/Fungal antagonist | Widespread | On numerous hosts. Soil, wood, numerous other substrates. | https://nt.ars-grin.gov/fungaldat/abases |

3. *Xylosandrus compactus* (Eichhoff)

Xylosandrus compactus (black twig borer or shot-hole borer) is a highly polyphagous pest of woody plants that probably originates from Asia and has been introduced to other parts of the world, most probably with the trade of plants and wood. It is widely distributed in Africa, Asia and South America. It has been introduced in the Pacific Islands, New Zealand, Southeastern USA, and more recently in Europe in Italy and Southern France (EPPO Alert-list, Rabaglia et al., 2006, Wood, 1982; Chong et al., 2009). It was first found in Europe in 2011 (Garonna et al., 2012) in urban parks of the Campania region of Italy. Then, the species has been recorded in Italy's Campania, Tuscany and Liguria, and recently emerged in South-east France. The first report in Europe of *X. compactus* and associated ambrosia fungi in a natural environment has been recorded in September 2016, in the Italian National Park Circeo, Central Italy, in the Latium Region (Vannini et.al., 2017).

The adult females are dark brown to almost shiny black, 1.4-1.9 mm long and about two times longer than wide. The small, wingless males are reddish black and measuring 0.9–1.3 mm in length (Hara & Beardsley, 1979). *Xylosandrus compactus* is a species in which males are born from unfertilized eggs (0.3 -0.5 mm) and females from fertilized ones. After mating, which primarily occurs between siblings just after adult emergence, the male remains in the gallery while the female leaves the tunnel through the entry hole and colonizes branches of new hosts, boring an entry hole and a subsequent brood gallery (Hara & Beardsley, 1979; Greco & Wright, 2015). (CABI Factsheet).

3.1. List of fungi isolated from *Xylosandrus compactus*

Table 2 shows the fungal species isolated from the *X. compactus* specimens in the Circeo Park (Italy). The fungal isolation was done according to the protocol described on **SAMFIX Deliverable: Fast routine protocol for detection of fungal symbiotic community associated with trapped Xylosandrus.**

Table 2. List of fungi isolated from *Xylosandrus compactus*

| SPECIES | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|--------------------------------|--------|--------------|-------------------------|-------------------------------|--------------|--|---|
| <i>Ambrosiella xylebori</i> | A | Microascales | abdomen, head, mycangia | Symbiont/plant pathogen | Cosmopolitan | Obligate, mutualistic symbionts of ambrosia beetles | Mayers et al., 2015 |
| <i>Acremonium camptosporum</i> | A | Hypocreales | abdomen, head, mycangia | Saprotrophs | Europe, Asia | dead plants or soil dwellers | Park, Thuong, Nguyen and Burm Lee, 2017 |
| <i>Acrodontium salmoneum</i> | A | Pleosporales | mycangia | Animal pathogen | Widespread | Wide-host range | Steiman et al., 1995 |
| <i>Alternaria alternata</i> | A | Pleosporales | abdomen, head, mycangia | Plant pathogens/Saprotrophs | Widespread | Wide-host range | Feng, Zheng, 2007 |
| <i>Aspergillus spelaeus</i> | A | Eurotiales | abdomen, mycangia | endophytes, food contaminants | Worldwide | soils and rhizospheres, indoor and cave environments | Hubka et al., 2017 |

| SPECIES | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|-------------------------------------|--------|-------------|-------------------------|-----------------------------|--|--|--|
| <i>Aspergillus flavus</i> | A | Eurotiales | abdomen | Plant pathogens/Saprotrophs | Cosmopolitan | Multitudinous substrates; secondary pathogen of some plants. Rots of fruit and food where it produces highly | Hubka et al., 2017 |
| <i>Aspergillus versicolor</i> | A | Eurotiales | head | Plant pathogens/Saprotrophs | Cosmopolitan | Numerous biological substrates. | Behnke-Borowczyk et al., 2019 |
| <i>Beauveria bassiana</i> | A | Hypocreales | abdomen | Animal pathogen | Cosmopolitan | Extremely wide host range; It can exist in diverse ecological environments including soil, plants and insects. | Imoulan et al., 2017 |
| <i>Beauveria pseudobassiana</i> | A | Hypocreales | external | Animal pathogen | Cosmopolitan | Wide insect host range; can survive in diverse environments. | Imoulan et al., 2017 |
| <i>Cladosporium cladosporioides</i> | A | Capnodiales | abdomen, head, mycangia | Saprotrophs/hyperparasitic | Cosmopolitan | Multiple genera in multiple families; Leaves, seeds, inflorescences, often dead plant material; Causing leaf spot and blight, flower blight, scab, sooty mold. | Barkat et al., 2016 |
| <i>Cladosporium perangustum</i> | A | Capnodiales | abdomen, head, mycangia | Saprotrophs | Widespread | Numerous hosts, associated with plants, fungi, food | Ogórek et al., 2012; Bensch et al., 2012 |
| <i>Cladosporium psychrotolerans</i> | A | Capnodiales | head, abdomen, mycangia | Saprotrophs | Europe, North America, Dominican Republic. | Hypersaline water in the Mediterranean basin, indoor and outdoor environment, occasionally from plants | Bensch et al., 2012 |

| SPECIES | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|-------------------------------------|--------|------------------|-------------------------|-----------------------------|-------------------------------|--|-------------------------------------|
| <i>Cladosporium ramontenellum</i> | A | Capnodiales | abdomen,head | Saprotrophs | South Africa, North America | Fruit and other plant material | K. Bensch et al., 2012 |
| <i>Cladosporium sphaerospermum.</i> | A | Capnodiales | head, abdomen | Saprotrophs | Cosmopolitan | Wide-host range; decaying Citrus leaves and branches in Italy; soil; decaying stem | Dugan et al. 2008; Zalaret al.,2007 |
| <i>Cladosporium uwebrauniana</i> | A | Capnodiales | head, mycangia | Unassigned | Europe | Indoor environment | Bensch et al., 2012 |
| <i>Clonostachys byssicola</i> | A | Hypocreales | abdomen, head, mycangia | Saprotrophs | Cosmopolitan | Fungi, plants | Alvandia and Hirooka, 2011 |
| <i>Clonostachys rosea</i> | A | Hypocreales | abdomen, head, external | Plant pathogens/Saprotrophs | Cosmopolitan | Various plant parts both living and newly killed,associated with bark beetle galleries | Nygren et al., 2018 |
| <i>Clypeosphaeria phillyreae</i> | A | Amphisphaeriales | abdomen | Unassigned | Europe | Phillyrea latifolia | Reblova, 2017 |
| <i>Cytospora acaciae</i> | A | Diaporthales | abdomen, head | Plant pathogen | worldwide | Broad-host range; Ceratonia siliqua in Spain | N. Jiang et al., 2020 |
| <i>Fomes fomentarius</i> | B | Polyporales | head | Saprotrophs | Temperate northern hemisphere | Broad host range; on dead or living hardwoods | Hashemi and Mohammadi, 2016. |
| <i>Fusarium solani</i> | A | Hypocreales | abdomen,head | Plant pathogen | Cosmopolitan | Broad host range; associated with ambrosia beetles | Sharma and Marques, 2018 |

| SPECIES | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|----------------------------------|--------|--------------|--------------------------|------------------|---|---|---|
| <i>Fusarium sporothichioides</i> | A | Hypocreales | head, mycangia, external | Plant pathogen | Temperate and tropical regions | Numerous hosts; found on roots, leaves, seeds, fruit causing root rot, leaf spot, dieback, etc | Arias et al., 2013; Taheri et al., 2017 |
| <i>Geosmithia flava</i> | A | Hypocreales | head, abdomen | Plant pathogen | Europe, North America (USA: CA), Europe, Asia and Australia | Numerous hosts; Beetle galleries, other plant substrates. In association with bark beetles | Kolarik et al., 2007 |
| <i>Geosmithia pallida</i> | A | Hypocreales | abdomen, head, mycangia | Plant pathogen | Widespread | Associated with bark beetle <i>Pseudopityophthorus pubipennis</i> ; Hosts: <i>Quercus</i> spp. (Fagaceae), <i>Prunus</i> spp., <i>Malus</i> (Rosaceae); Causing Foamy bark canker, dieback, death | Kolarik et al., 2017 |
| <i>Geosmithia</i> sp. 21 NL-2015 | A | Hypocreales | abdomen, head | Unassigned | California | Wide range of host plants found in association with different bark and ambrosia beetles | Kolarik et al., 2017 |
| <i>Paraphoma fimeti</i> | A | Pleosporales | mycangia | Saprotrophs | Cosmopolitan | Soil, dead plant tissues; Herbaceous and woody plants; roots of <i>Juniperus communis</i> | Boeremaet al., 2004; De Gruyter et al., 2010; Moslemiet al., 2017 |
| <i>Penicillium citrinum</i> | A | Eurotiales | head | Saprotrophs | Cosmopolitan | Soil, decaying vegetation, variety of organic substrates | Kozakiewicz, 1992 |

| SPECIES | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|-----------------------------------|--------|------------|--------------|-----------------------------|-------------------|---|---|
| <i>Penicillium coccotrypicola</i> | A | Eurotiales | abdomen | Saprotrophs | Australia | Galleries of palm seed borer <i>Coccotrypes carpophagus</i> ; <i>Archontophoenix cunninghamiana</i> (Arecaceae) | Crous et al., 2014 |
| <i>Penicillium glabrum</i> | A | Eurotiales | external | Saprotrophs | Cosmopolitan | Numerous substrates; Numerous hosts; causing post harvest fruit rot | Duduk et al., 2017 |
| <i>Penicillium glabrum</i> | A | Eurotiales | mycangia | Saprotrophs | Cosmopolitan | Numerous substrates; Numerous hosts; causing post harvest fruit rot | Duduk et al., 2017 |
| <i>Penicillium multicolor</i> | A | Eurotiales | abdomen,head | Plant pathogens/Saprotrophs | Asia; Europe; USA | Leaves, seeds, coniferus and broadleaved sp. | Visagie et al., 2013 |
| <i>Penicillium pancosmium</i> | A | Eurotiales | abdomen | Plant pathogens/Saprotrophs | Worldwide | on hardwood log, Isolated from soil, old <i>Armillaria mellea</i> on a hardwood log, <i>Piptoporus</i> (on <i>Betula</i> sp), nut of <i>Juglans cinerea</i> (butternut) and porcupine dung. | Houbraken et al.,2015 |
| <i>Peniophora meridionalis</i> | B | Russulales | external | Plant pathogen | Europe | Wood; deciduous species; <i>Ceratonia siliqua</i> , <i>Erica</i> sp. <i>Eucalyptus</i> sp., <i>Q. Ilex</i> , <i>Q. Pyrenaica</i> , <i>Pistacia lentiscus</i> , <i>P. Halepensis</i> , <i>Arbutus unedo</i> , <i>Castanea</i> sp., <i>Viburnum tinus</i> | https://www.gbif.org/species/2552347 |

| SPECIES | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|----------------------------------|--------|----------------|-----------|--|--------------------------|---|---|
| <i>Pestalotiopsis biciliata</i> | A | Xylariales | external | Plant pathogens/Saprotrophs | Cosmopolitan | Within woody species, both conifers and broadleaf trees are reported as hosts.causing a various range of symptoms including leaf spots, leaf blight, fruits rot as well as post-harvest diseases. | Morales-Rodríguez et al., 2018 |
| <i>Peziza ostracoderma</i> | A | Pezizales | mycangia | Saprotrophs | North America and Europe | Peat mold | Lohr et al., 2017 |
| <i>Phanerochaete livescens</i> | B | Polyporales | mycangia | Saprotrophs | Widespread | Alnus glutinosa, A. incana, A. hirsuta, Quercus sp., Fagus sylvatica, Populus tremula, Corylus avellana, Acer platanoides, Padus avium | Volubev et al., 2015 |
| <i>Sarocladium strictum</i> | A | Hypocreales | external | Plant pathogen | Cosmopolitan | Broad host range, associated with ambrosia beetle Eucallitricus formicatus | Farr and Rossman, 2020; Li et al., 2016 |
| <i>Simplicillium lamellicola</i> | A | Hypocreales | head | plant-parasitic, symbiotic, entomopathogenic | Widespread | broad spectrum of hosts and substrates, such as insects, plants, rusts, nematodes and mushrooms | De-Ping et al., 2019 |
| <i>Sistotrema brinkmannii</i> | B | Cantharellales | abdomen | Saprotrophs | Widespread | Usually on wood, sometimes on plant debris and basidiomata | https://nt.ars-grin.gov/fungaldatabases |

| SPECIES | PHYLUM | ORDER | BODY PART | FUNCTIONAL GUILD | OCCURANCE | REPORTED HOST/SUBSTRATE | REFERENCE |
|---------------------------------|--------|----------------|-------------------------------|-------------------------------|-----------------------------------|---|---|
| <i>Talaromyces amestolkiae</i> | A | Eurotiales | head, abdomen, mycangium, ext | Animal pathogen | Cosmopolitan | Talaromyces contains species that are medically important. Emerging pathogen of agricultural crops. | Tsang et al., 2017; Yilmaz et al., 2014 |
| <i>Talaromyces purpurogenus</i> | A | Eurotiales | head | Animal pathogen | Cosmopolitan | Talaromyces contains species that are medically important. Emerging pathogen of agricultural crops | Tsang et al., 2017; Yilmaz et al., 2014 |
| <i>Torrubiella alba</i> | A | Hypocreales | head, mycangia | Animal pathogen | Cosmopolitan | Obligate symbiont with plants, animals and other fungal species | Johnson et al., 2008 |
| <i>Trichoderma hamatum</i> | A | Hypocreales | head, abdomen mycangium, ext | Saprotrophs/Fungal antagonist | Cosmopolitan | Found on roots and other plant parts on numerous hosts; causing soft roots | Bissett et al., 2015; Han et al., 2017 |
| <i>Umbelopsis westeae</i> | M | Mucorales | abdomen | Saprotroph | Australia | Different soil substrates | Wang et al., 2013 |
| <i>Ustilaginoidea virens</i> | A | Incertae sedis | abdomen | Plant pathogen | Worldwide in rice-growing regions | Substrate: Inflorescence/inflorescence; Host: <i>Oryza sativa</i> , <i>Zeamays</i> , <i>Brachiariabrizantha</i> (Poaceae). | Kumari and Sharma, 2017; Fan et al., 2016 |
| <i>Xenoacremonium falcatus</i> | A | Hypocreales | head | Unassigned | Asia, Europe | <i>Castanea sativa</i> ; other substrates | Aghyeva et al., 2017 |

4. CONCLUSION

Fungi typically live in highly diverse communities composed of multiple ecological guilds. "Plant-pathogens" have been the most abundant fungal-guild present on the isolated fungi from *Xylosandrus crassiusculus* and *X. compactus*. Of particular interest is the finding of several plant pathogens associated to different parts of the insect and having as hosts the family of Fagaceae or other tree species. For example: *Cryphonectria parasitica*, the causal agent of chestnut blight, isolated from *X. crassiusculus* or *Pestalopsis biciliata*, the causal agent leaf blotch symptoms on Eucalyptus. The genus *Fusarium* and the species complex *Fusarium solani* (FSSC) were isolated from both *Xylosandrus* species. *Fusarium solani* it is a symbiotic fungus cultivated in tunnels of host plants by the female pest, which is attracted to volatiles from *F. solani* (Egonyu, 2017). Furthermore, members of genus *Fusarium* have been reported in association with other ambrosia beetles, and they are often reported as pathogenic to the host tree and other woody crops (i.e., avocado) in Sicily (Gugliuzzo *et al.*, 2020). Species belonging to the genus *Fusarium* have diverse ecological functions as they can act as saprophytes, endophytes and animal and plant pathogens. It is important to mention that the genus *Fusarium* includes important plant pathogens that affect both forest and agricultural species by producing different types of wall-degrading enzymes (e.g., cellulases, glucanases and glucosidases) and mycotoxins such as beauvericin and fumonisins (Bezoz, 2018; Sharma, 2018). *Geosmithia pallida* is a species native to Europe (Lynch *et al.*, 2014) and one of the most diffused fungal species in Mediterranean maquis, closely associated with alien species. *G. pallida* appear to be more a no specific commensal. It was reported from other plant-insect interactions, such as *Castanea sativa* and the Cynipidae wasp *Dryocosmus kuriphilus* (Morales-Rodríguez *et al.*, 2019), *Carya illinoensis* and *Quercus laurifolia* with *Pseudopityophthorus minutissimus* (Huang *et al.*, 2019) or associated with *X. compactus* at the National Park of Circeo (Vannini *et al.*, 2017). But it can also behave as a plant pathogen, for instance, *G. pallida* have been reported in the literature as a causal agent of foamy bark canker in *Quercus agrifolia* in Californian association with *Pseudopityophthorus pubipennis* (Lynch *et al.*, 2014). According to the literature, *G. pallida* was accidentally introduced from Europe, like an alien pathogen of live oaks in the United States (Lynch *et al.*, 2014).

The introduction of ambrosia beetle *Xylosandrus* and consequently fungal species, which represent a prevalent group of forest pathogens, as they are the major component of biodiversity in Europe and second-largest group of Eucaryotes right after insects. Many fungal species are considered as cryptogenic, which means they are most likely alien but with unknown origin, as they are poorly represented in alien species databases and unfortunately there is poor knowledge of their biogeography. For example, species as *Paraconiothyrium archidendri* reported on Asia, *Acremonium roseolum* on Asia and South America or *Geosmithia* sp. 21 on USA; wich roles should be more investigated.

Symbiosis plays a critical role when the insects attempt to invade a new habitat. Understanding the ecological factors that influence the adaptation of an organism in a new environment and the uptake of new microorganisms are the key to explain the mechanism of biological invasions. Right one of the most complex examples of symbiosis is the one between ambrosia beetles and ambrosia fungi. Nevertheless, should be considered that different kind of fungal species are associated to different species of ambrosia beetles and this is the reason why some of the fungal species associated to some beetles are not found in association with *X. compactus/crassiusculus* or vice versa, as there is the difference between fungal species that are associated to an exotic or native ambrosia beetle species. After the introduction of an exotic species such as *Xylosandrus*, in a new environment, there is a series of biotic and abiotic forces that greatly influence the community of organisms in association with the insect. It is considered that forest habitat strongly influences the diversity of fungal species associated with the exotic ambrosia beetles. The absence of adaptation of exotic species could limit its establishment in a new environment. However, gaining microorganisms native to the invaded environment may support the exotic species to overcome these ecological barriers (Rassati et al., 2019).

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